

Protocol of HEK293-TRHR Cell-based Assay for High-throughput Screening

DOCUMENT: HEK293-TRHR_TOX21_SLP_Version1.0
TITLE: Protocol of HEK293-TRHR Cell-based Assay for High-throughput Screening

ASSAY REFERENCES:

Assay Target	Cell Lines	Species	Tissue of Origin	Assay Readout	Assay Provider	Toxicity Pathway
TRHR	Hek293-TRHR	Human	Embryonic kidney	Fluorescence	Codex	Gs-coupled TRHR signaling

QUALITY CONTROL PRECAUTIONS:

1. The cells should not be grown more than 80-85% confluence
2. Do not leave cells in Trypsin for more than 5 min at RT

MATERIALS and INSTRUMENTS:

Supplies/Medium/Reagent	Manufacturer	Vender/Catalog Number
DMEM, high-glucose	Invitrogen	11995
FBS	Invitrogen	26140
Penicillin/Streptomycin (antibiotic)	Invitrogen	15140
Puromycin	Invitrogen	A11138
Recovery Cell Culture Freezing Medium	Invitrogen	12648
Cal-520 calcium assay kit	AAT Bioquest	36400
TRH	Sigma	P1319
FDSS 7000EX kinetic plate reader	FDSS 7000EX	-
Multidrop	Thermofisher	-
BioRAPTR	Beckman Coulter	-

PROCEDURE:

1. Cell handling:
 - 1.1. Media Required:

Component	Growth Medium	Assay Medium	Thaw Medium	Freezing Medium
DMEM, high glucose	90%	90%	90%	-
FBS	10%	10%	10%	-
Penicillin/Streptomycin (antibiotic)	1%	1%	1%	-
Puromycin	1ug/mL	1ug/mL	-	-
Recovery Cell Culture Freezing Medium	-	-	-	100%

1.2. Thawing method

- 1.2.1 Place 14 mL of pre-warmed thaw medium into a 15 mL conical tube
- 1.2.2 Remove the vial of cells to be thawed from liquid nitrogen and thaw rapidly by placing at 37°C in a water bath
- 1.2.3 Mix the entire content of the vial to 14 ml of pre-warmed medium and centrifuge to remove DMSO
- 1.2.4 Discard the supernatant and reconstitute the pellet using pre-warmed thaw medium
- 1.2.5 Transfer the cells to T75 flask

1.3. Propagation method

- 1.3.1 Detach the cells from the flask using Trypsin-EDTA (0.05%)
- 1.3.2 The cells in growth medium are re-seeded in T225 flask

2. Assay Protocol

- 2.1 Harvest cells and re-suspend in assay medium and adjust the required cell density
- 2.2 Dispense 1500 cells/4 μ L/well into 1536 well tissue treated black-clear-bottom plates using a Multidrop dispenser
- 2.3 Incubate the plates at a 37°C, 5% CO₂ incubator for 18 h
- 2.4 Add 4 μ L of Loading Buffer
- 2.5 Incubate at 37°C, 5% CO₂ for 2 h
- 2.6 Incubate at RT for 15 min
- 2.7 Transfer 23nL of compounds and positive control to the assay plate by a pin tool.
- 2.8 Read the fluorescence intensity in FDSS 7000EX kinetic plate reader with a filter set of Ex/Em=480/540 for 3 min at 1 sec intervals
- 2.9 For antagonist mode, add 1 μ L of TRH at 0.1 nM (final) to each well except 16 wells in column 4 after 5 min incubation at RT.
- 3.0 Read the fluorescence intensity in FDSS 7000EX kinetic plate reader with a filter set of Ex/Em=480/540 for 3 min at 1 sec intervals

3. Assay Performance

TRHR-agonist (TSH)	Online Validation (Mean \pm SD)
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AC50 (nM)	0.05±0.04
S/B	48.15 ± 12.93
CV (%)	25.90 ± 8.17*
Z'	0.79 ± 0.06
N	27

TRHR-antagonist (N/A)	Online Validation (Mean ± SD)
AC50 (nM)	N/A
S/B	62.58 ± 11.05
CV (%)	6.70 ± 1.32*
Z'	0.81 ± 0.05
N	27

*CV values shown represent average of DMSO plates and low concentration plates only (n=18).