

Protocol of AR-MDA MDA-MB-453 Cell-based Assay for High-throughput Screening

DOCUMENT: AR-MDA_TOX21_SLP_Version1.0

TITLE: Protocol of AR-MDA MDA-MB-453 Cell-based Assay for High-throughput Screening

ASSAY REFERENCES:

| Assay Target | Cell Lines | Species | Tissue of Origin | Assay Readout | Assay Provider | Toxicity Pathway |
|-------------------|------------|---------|-----------------------|---------------|----------------|------------------|
| Androgen receptor | MDA-MB-453 | Human | Mammary gland, breast | Luminescence | ATCC | NR signaling |

QUALITY CONTROL PRECAUTIONS:

1. -Maintain cell culture below 85-90% confluence
2. -Cell culturing and assay culture doesnot require CO2

MATERIALS and INSTRUMENTS:

| Supplies/Medium/Reagent | Manufacturer | Vender/Catalog Number |
|--|------------------------------|------------------------------|
| -Leibovitz's L-15 Medium | -ATCC | -ATCC / 30-2008 |
| -Fetal Bovine Serum | -Hyclone | -Hyclone / SH30071.03 |
| -Penicillin/Streptomycin | -Invitrogen | -Invitrogen / 15140 |
| -Recovery Cell culutre Freezing Medium | -Invitrogen | -Invitrogen / 12648 |
| -0.25% Trypsin-EDTA | -Invitrogen | -Invitrogen / 25200 |
| -1536-well white solid plates | -Greiner Bio-One | -Greiner Bio-One / 789173-F |
| -MULTIDROP COMBI | -Thermo Electron Corporation | -Thermo Electron Corporation |
| -BioRAPTR FRD | -Beckman Coulter | -Beckman Coulter |
| -ViewLux Plate Reader | -Perkin Elmer | -Perkin Elmer |

| | | |
|---|---------------|-----------------------------|
| -ONE-Glo(TM) Luciferase Assay System | -Promega | -Promega / E6120 |
| -CellTiter-Fluor(TM) Cell Viability Assay | -Promega | -Promega / G6082 |
| -R1881 (Agonist Reference Compound) | -Perkin Elmer | -Perkin Elmer / NLP005005MG |
| -Nilutamide (Antagonist Reference Compound) | -Sigma | -Sigma / N8534 |
| -Tetraoctyl ammonium bromide (Viability Reference Compound) | -Sigma | -Sigma / 294136 |

PROCEDURE:

1. Cell handling:

1.1. Media Required:

| Component | Growth Medium | Assay Medium | Thaw Medium | Freezing Medium |
|--|---------------------|---------------------|---------------------|-----------------|
| -Leibovitz's L-15 Medium | -100% | -100% | -100% | - |
| -Fetal Bovine Serum | -10% | -10% | -10% | - |
| -Penicillin & Streptomycin | -100U/ml & 100ug/ml | -100U/ml & 100ug/ml | -100U/ml & 100ug/ml | - |
| -Recovery Cell culture Freezing Medium | - | - | - | -100% |

1.2. Thawing method

1.2.1 -Thaw a vial of cells in 9ml of pre-warmed thaw/culture medium and then centrifuge

1.2.2 -Resuspend the pellet with the thaw/culture medium and seed at 2 million cells per T-75 flask

1.3. Propagation method

1.3.1 -Trypsinize cells from the culturing flask and centrifuge and then resuspend cells in culture medium

1.3.2 -Passage cells at 6-7 million per T-225 flask

2. Assay Protocol

2.1 -Trypsinize cells from the culturing flask and centrifuge and then resuspend cells in culture/assay medium

2.2 -Dispense 3000 cells/5uL/well (for agonist mode) into 1536-well tissue treated white/solid bottom plates using a 8 tip dispenser (Multidrop)

2.3 -Incubate the assay plates for 5-6hrs at 37C and 0% CO2

2.4 -Transfer 23nL of compounds from the library collection and positive control/DMSO into the assay plates through Pintool

2.5 -Compound transfer was followed by the addition of 1uL of 0.5nM (final concentration) R1881 or assay buffer using 2 tips of a dispenser (BioRAPTR FRD)

- 2.6 -Incubate the assay plates for 16hrs at 37C and 0% CO₂
- 2.7 -After 15hrs of incubation at 37C and 0%CO₂, 1ul of CellTiter-Fluor reagent was added using a single tip of a dispenser (BioRAPTR FRD)
- 2.8 -Incubate the assay plates at 37C and 0%CO₂ for 1hr
- 2.9 -Read fluorescence using ViewLux plate reader
- 2.10 -Then followed by the addition of 4ul of ONE-Glo(TM) Luciferase reagent using a single tip of a dispenser (BioRAPTR FRD)
- 2.11 -Incubate the assay plates at room temperature for 30min
- 2.12 -Read Luminescence using ViewLux plate reader